### Anatomy

1	Course name	Anatomy
2	Course Code	PH 108
3	Course type: /general/specialty/optional	General
4	Accredited units	1 credits
5	Educational hours	1 hours/week/THEORY
6	Pre-requisite requirements	Biology
7	Program offered the course	Department of Biomedical Science
8	Instruction Language	English
9	Date of course approval	12/2021

Brief Description: This subject is designed to impart fundamental knowledge on the s and functions of the various systems of the human body, such as cardiovascular, endocrine systems. Also the subject provides the knowledge on the structure and functions of the respiratory, of urinary and reproductive systems. It deals with structure and func- and lymphatic, skeletal and muscular systems. The subject prov- basic knowledge required to understand the various discip pharmacy.	
Textbooks required for this Course:	1- Principles of Anatomy and Physiology by Tortora Grabowski. Palmetto, GA, U.S.A.
Course Duration	28 weeks
Delivery	Lecture-based, Group interaction and discussion, Self-directed activities, Active participation. Lectures (Tools: board, data show, models and discussion). The lectures were added on the internet site of the faculty to be available to the students all the time as learning.
Course Objectives:	<ul> <li>Upon completion of this course the student should be able to</li> <li>1. Explain the gross morphology, structure and functions of various organs of the humanbody.</li> <li>2. Describe the various homeostatic mechanisms and their imbalances.</li> <li>3. Identify the various tissues and organs of different systems of human body.</li> <li>5. Appreciate coordinated working pattern of different organs of each</li> </ul>

	system		
Course Assessments	Miduear evam	20%	
Course Assessments		20%	
	- Final theoretical exam	60%	
	- Total	100%	
Content Breakdown	Content Breakdown Topical Coverage		
Topical Coverage	· -		
Session 1 (Week 1)	Unit I: Nervous system:		
	Sections of the nervous system, the most impo	rtant structural and	
	functional differences of the sympathetic ar	nd parasympathetic	
	nerves		
	-Spinal-cerebral nerves		
Session 2 (Week 2)	Physiology of nerve impulses and reflex arc     Corobrogonical fluid, its composition, charactericity	tice avala functions	
	Cerebrospinal nuiu, its composition, characterist medical importance	ICS, CYCle, Tunctions,	
	Some neurological diseases (cerebral and	vascular accidents.	
	Parkinson's disease	vuscului ucc,	
Session 3 (Week 3)	Unit II: Cardiovascular system:		
	Its components and most important functions		
	Hemoglobin in terms of its composition, types, characteristics, and		
	importance Blood clot and its formation steps		
	<ul> <li>Blood clot and its formation steps</li> <li>Blood groups and their medical importance</li> </ul>		
	<ul> <li>Blood groups and their medical importance</li> <li>Anemia and its types</li> </ul>		
	Anemia and its types.		
Session 4 (week 4)	Ine most important anatomical features of the heart and its     function		
	<ul> <li>Heart cycle, heart capacity and factors affecting it</li> </ul>		
	<ul> <li>Circulation and the most important changes that occur to the</li> </ul>		
	circulatory system before and after birth.		
Session 5 (Week 5)	Unit III: Endocrine system:		
	Definition of hormones and their types		
	• The mechanism of the effect of hormones	on tissues and the	
	relationship of primary and secondary messengers to receptors		
	How to control the secretion of hormones?		
Session 6 (Week 6)	<ul> <li>Study of the following glands from an anator their corrections, the functions of these horms</li> </ul>	nical point of view,	
	their secretions, the functions of these hormonal secretions, and		
	The nituitary gland, pineal gland, thyroid and	parathyroid glands.	
	thymus gland, parathyroid glands, pancreas, te	stes and ovaries are	
	studied within the reproductive system.		
Session 7 (Week 7)	Unit IV: Urinary system:		
	<ul> <li>Its parts and functions</li> </ul>		
	<ul> <li>How urine is formed in the renal tubules and the</li> </ul>	e urea cycle	

Session 8 (Week 8)	Unit V: Reproductive system:	
	<ul> <li>Its parts (male and female and its general functions)</li> </ul>	
	<ul> <li>Steps in the formation of sperm and eggs</li> </ul>	
	<ul> <li>Menstrual cycle, fertilization, pregnancy and how to regulate it</li> </ul>	
	Fetal development	
Session 9 (Week 9)	Unit VI: Respiratory system:	
	<ul> <li>Its parts, most important characteristics and functions</li> </ul>	
	<ul> <li>Respiratory volumes and lung capacity</li> </ul>	
	Inhale and exhale steps	
	• Steps for the transfer of gases (O2, CO2) between tissues and lungs	
	and the factors affecting the saturation of blood with gases and	
	then poisoning by CO.	
Session 10 (Week 10)	Regulating breathing (control centers of the nervous system)	
	<ul> <li>Adaptation of the body to the change in altitude (diving and alighbrance)</li> </ul>	
Session 11 (Meak 11)	climbing to high areas)	
Session 12 (Week 11)		
Session 12 (Week 12)	Midyear exam	
Session 15 (Week 15)		
Session 14 (Week 14)	Linit VIII: Digostivo system:	
Session 12 (Week 15)	• Its parts and general functions	
	<ul> <li>The most important changes that occur to carbohydrates, fats and</li> </ul>	
	<ul> <li>The most important changes that occur to carbonydrates, rats and protein as a result of digestion in the alimentary canal</li> </ul>	
Session 16 (Week 16)	The role of the annendices of the alimentary canal (liver and	
56551011 10 (Week 10)	nancreas) in the digestive process	
Session 17 (Week 17)	Absorption of indigestible substances	
	<ul> <li>Metabolism and energy release (anaerobic and aerobic respiration)</li> </ul>	
Session 18 (Week 18)	Unit VIII: Lymphatic system:	
	<ul> <li>Its parts, its role, and its importance</li> </ul>	
	<ul> <li>Specialized and non-specialized immune systems, cellular and</li> </ul>	
	humoral immunity	
Session 19 (Week 19)	Unit IX:Integumentary system:	
	Structure and function	
Session 20 (Week 20)	Unit X: the skeletal system:	
	• The most important features of bones, their general functions, and	
	their medical importance	
	Distinctive superficial signs of bone.	
	Bone shapes (types).	
Session 21 (Week 21)	• Study of the spine in terms of the intervertebral discs (their	
	structure and function), as well as the most important superficial	
	anatomical differences for the vertebrae of the five regions of the	
	spine.	
	The most important differences between the male and female     structure	
Session 22 (Maak 22)	Studuing the douglapment of the middle car Oscioles in humans and	
Jession ZZ (Week ZZ)	<ul> <li>Studying the development of the middle ear Ossicles in humans and comparing them with other vertebrates</li> </ul>	
	ioints	

	Factors affecting bone growth		
	• Brief indication of some diseases of the skeletal system (disc		
	herniation, kyphosis and lordosis)		
Session 23 (Week 23)	Unit XI: the muscular system:		
	<ul> <li>The most important differences between smooth, skeletal, and</li> </ul>		
	cardiac muscles		
	General muscle functions		
	<ul> <li>Physiology of muscle contraction, sliding filament theory and its</li> </ul>		
	comparison with smooth muscle contraction		
Session 24 (Week 24)	Cardiac muscle as a conduction organ		
	<ul> <li>Muscular fatigue and muscle energy sources</li> </ul>		
	Muscle cramps		
	<ul> <li>Some muscle diseases (myalgia and myasthenia gravis)</li> </ul>		
Session 25 (Week 25)	Review		
Session 26 (Week 26)	Review		
Session 27 (Week 27)			
Session 28 (Week 28)	Veek 28)		
Final theoretical exam			
Attendance	Students are expected to attend every session of class, arriving on time,		
Expectations	returning from breaks promptly and remaining until class is dismissed.		
	Absences are permitted only for medical reasons and must be supported		
	with a doctor's note.		
Generic Skills	The faculty is committed to ensuring that students have the full range of		
	knowledge and skills required for full participation in all aspects of their		
	lives, including skills enabling them to be life-long learners. To ensure		
	graduates have this preparation, such generic skills as literacy and numeric,		
	computer, interpersonal communications, and critical thinking skills will be		
	embedded in all courses.		
Course Change	Information contained in this course outline is correct at the time of		
	publication. Content of the courses is revised on an ongoing basis to ensure		
	relevance to changing educational employment and marketing needs. The		
	relevance to changing educational employment and marketing needs. The		
	relevance to changing educational employment and marketing needs. The instructor will endeavor to provide notice of changes to students as soon as		

# Histology

1	Course name	Histology
2	Course Code	PH107
3	Course type: /general/specialty/optional	General
4	Accredited units	2 credits (1 lecture + 1 lab / week)
5	Educational hours	1 hour/week/Theory, 2 hours/ week/ lab
6	Pre-requisite requirements	Biology
7	Program offered the course	Biomedical science department
8	Instruction Language	English
9	Date of course approval	12/2021

Brief Description:	Histology: In this course the student will study the technique of microscopic	
	use, the basic and general introduction to histology of the following systems:	
	- Cell, types of tissues, CVS, GIT, genitourinary systems, respiratory system,	
	immune system, endocrine and male and female systems.	
Textbooks required for this Course:	<ol> <li>Junqueira, L. C. and J. Carneiro, Basic Histology, tenth edition. Norwalk: Appleton &amp; Lang; Last edition.</li> <li>Eroschenko, V. P., di Fiore's Atlas of Histology with Functional Correlations, ninth edition. Philadelphia: Lippincott Williams &amp; Wilkins; Last edition.</li> <li>Text Basic Histology by Luiz Carlos 11th ed. (2005)</li> </ol>	
Course Duration	28 weeks	
Delivery	<ul> <li>Lectures (Tools: board, data show).</li> <li>Tutorials and group discussions.</li> <li>Assignments (if applicable).</li> <li>Videos.</li> <li>Practical classes (Lab experiments+ computerized experiments simulation).</li> </ul>	
Course Objectives:	1. To understanding the technique in use of microscope.	

	5- State the functions of the exocrine and endocrine glands.		
Course Assessments	Midyear exam	20%	
	Quizzes, reports, presentation	10%	
	Practical continuous assessment, exam	10%	
	Final Practical exam	20%	
	Final theoretical exam	40%	
	Total	100%	
Content Breakdown	Content Breakdown Topical Coverage		
Topical Coverage			
Session 1 (Week 1)	Unit I: Introduction and methods of study:		
	- Microscopes and micro technique		
Session 2 (Week 2)	Unit II: Cytology: I- Cell structure:		
	a) Cytoplasm.		
	b) Organoids		
Session 3 (Week 3)	c) Cytoskeleton.		
Session 4 (Week 4)	II- Cell division and cell death .		
Session 5 (Week 5)	Unit III: Epithelial tissues:		
	A- General characteristics.		
	B- Classification of epithelia:		
Session 6 (Week 6)	C- Surface modifications and cell junctions.		
	D- Functions of epithelial tissues.		
Session 7 (Week 7)	Unit IV: Connective tissues:		
	A- General characteristics and classification.		
	B- Cells, ground substance and fibers.		
Session 8 (Week 8)	Unit V: Cartilages: General characteristics and classification.		
Session 9 (Week 9)	Unit VI: Bones:		
	A- General characteristics and classification		
Session 10 (Week 10)	B- Cell and matrix		
Session 11 (Meak 11)	C- Structure of compact and spongy bones.		
Session 12 (Week 11)			
Session 13 (Week 13)	Midyear exam		
Session 14 (Week 14)	-		
Session 15 (Week 15)	Unit VII: Blood:		
	A- General structure (cells, plasma, and stains)		
Session 16 (Week 16)	B- Erythrocytes, leukocytes, and blood platelets		
Session 17 (Week 17)	C- Bone marrow		
Session 18 (Week 18)	Unit VIII: Muscular tissues:		
	A- General characteristics and classification		
Session 19 (Week 19)	B- Skeletal, smooth, and cardiac muscles		

Session 20 (Week 20)	Unit IX: Nervous tissue:	
	A- General structure.	
	B- Neurons.	
	C- Neuroglia.	
	D- Nerve trunk and ganglia	
Session 21 (Week 21)	Unit X: Cardiovascular system:	
	Cardiovascular system:	
	$\Delta_{-}$ General structure of the blood vessels	
	B- structure of the arteries veins and canillaries	
Session 22 (Week 22)	Unit XI: Lymphatic system:	
	A- Immune system.	
	B- Lymph node, spleen, thymus gland, and tonsil	
Session 23 (Week 23)	Unit XII: Integumentary system (skin and its appendages):	
	A- General structure	
	B- Appendages and glands	
	C- Functions of skin	
Session 24 (Week 24)	Unit XIII: Respiratory system	
	A- Conducting portion (Nose, Nasonhanyny, Trachea Bronchus & Bronchiolos)	
	A- Conducting portion (Nose, Nasopharynx, Hachea Dionenus & Dionenuses)	
Session 25 (Week 25)	Init XIV. Digostivo system	
Jession 25 (Week 25)		
	A- General structure of esophagus, stomach, small and large intestine,	
	rectum, anus, liver, and pancreas.	
	B- Glands associated with the digestive tract (Salivary glands, Pancreas, Liver	
	& Gall bladder.	
Session 26 (Week 26)	Unit XV: Urinary system	
	A- Structure & Function of the (kidney &nephron)	
	B -Histology of the nephron (filtration, absorption & excretion).	
	C - Structure of the (Ureter, Bladder & Urethra).	
Session 27 (Week 27)	Unit XVI: Endocrine system	
	A -General structure of the Pituitary gland Adrenal Thyroid Parathyroid	
	A -General structure of the Pitultary gland, Adrenal, Inyroid, Parathyroid,	
Session 28 (Week 28)	Unit XVI:	
	Male reproductive system:	
	A -Excretory genital ducts-Excretory genital glands (Seminal vesicles, Prostate	
	& Cowper'sglands)	
	Female reproductive system:	
	A-General structure of ovary, Oviduct, Uterus & Vagina.	
	B -Stages of follicle developmentOvulation	
	Final theoretical exam	
Practical work	PART II· PRACTICAL Histology (One lab / week)	
	1. Light Microscope	
	2. EM of cell organelles	
	3. Epithelium	
	4. Connective tissues	
	5. Cartilage	
	0. Build	

	<ul> <li>8. Muscles</li> <li>9. Neural tissue</li> <li>10. Blood vessels</li> <li>11. Lymphatic tissue</li> <li>12. Spleen</li> <li>13. Digestive system</li> <li>14. Skin</li> </ul>	
	Final Practical Exam	
Attendance Expectations	Students are expected to attend every session of class, arriving on time, returning from breaks promptly and remaining until class is dismissed. Absences are permitted only for medical reasons and must be supported with a doctor's note.	
Generic Skills	The faculty is committed to ensuring that students have the full range of knowledge and skills required for full participation in all aspects of their lives, including skills enabling them to be life-long learners. To ensure graduates have this preparation, such generic skills as literacy and numeric, computer, interpersonal communications, and critical thinking skills will be embedded in all courses.	
Course Change	Information contained in this course outline is correct at the time of publication. Content of the courses is revised on an ongoing basis to ensure relevance to changing educational employment and marketing needs. The instructor will endeavor to provide notice of changes to students as soon as possible. Timetable may also be revised.	

#### Biochemistry

1	Course name	Biochemistry
2	Course Code	РН 202
3	Course type: /general/specialty/optional	General
4	Accredited units	3 units (2 hrs./week theoretical
		2 hrs./week practical)
5	Educational hours	4 hours / week
6	Pre-requisite requirements	Biology and organic chemistry
7	Program offered the course	Department of Pharmaceutical chemistry
8	Instruction Language	English
9	Date of course approval	12/2021

<b>Brief Description:</b>	The course is designed to give students the important foundations of		
•	biochemistry. This course focuses on processes occurring at a molecular		
	level. It focuses on what's happening inside human cells, studying		
	components such as carbohydrates, li	pids, proteins, and organelles. Also the	
	course covers enzymes, haem metak	polism, vitamins, and nucleotides and	
	nucleic acids. In addition, it includes	practical part (preparation of buffers,	
	quantitative and qualitative test of the	e above topics)	
Textbooks required	1. Marks' Essentials of Medical Bi	ochemistry A Clinical Approach. By	
for this Course:	Michael Lieberman and Alisa Peet. M	ichael Tully.	
	2. Practical Textbook of biochemistry	for medical students. By	
	Divivasudevan and Subir Kumar Das. J	aypeebrother's medical publishers.	
	A Biochemistry Research Internationa	l www.hindawi.com/journals/hri	
Course Duration	28 weeks	www.initdawi.com/jodmais/bit	
Course Duration	- Lectures (Tools: board_data show)		
	-Tutorials and group discussions.		
	-Assignments (if applicable), seminar	s, researches and posters.	
Delivery	-Videos.	_	
	- Practical classes (Lab experiments+ c	computerized experiments simulation).	
	• The lectures are added on the interr	net site of the faculty to be available to	
	the students all the time as an <i>e</i> -learning.		
Course Objectives:	By the end of the course, students should be able to:		
<b>Course Assessments</b>	Midyear exam	20%	
	Quizzes, reports, presentation	10%	
	Practical continuous assessment,	10%	
	exam		
	Final Practical exam	20%	
	Final theoretical exam	40%	
	Total	100%	
Content Breakdown Topical Coverage	Content Breakdown Topical Coverage		
Session 1 (Week 1)	Properties of water and buffers:		
	Water and acid base balance.		
	Buffer, acidosis, and alkalosis		
Session 2 (Week 2)	Chemistry of carbohydrates:		
	Definition and classification of carboh	ydrates.	
	Different types of complex carbohydrates.		
Session 3 (Week 3)	Carbohydrate metabolism:		
	- Pentose phosphate pathway, its importance, deficiency of G6PDs		
Session 4 (Mook 4)		stances aluconeogenesis	
Jession 4 (Week 4)	- Giuconeogenesis: giuconeogenic substances, gluconeogenesis		
Session 5 (Week 5)	- Tricarboxylic acid cycle: It's reactions functions role in metabolism and		
	ATP production by respiratory chain and regulation		
Session 6 (Week 6)	Chemistry of lipids:		
	-Lipid classification		
	-Physical properties		
	- Reactions of fatty acid		

Session 7 (Week 7)	-Types of lipid:	
	i- phospholipids,	
	ii- glycolipids	
	iii- lipoproteins	
	iv- triacylglycerol	
	v- cholesterol	
	-Lipoproteins	
Session 8 (Week 8)	Lipid metabolism:	
	Biosynthesis of fatty acids and Ketone bodies	
Session 9 (Week 9)	Biosynthesis and catabolism of triglycerides, phospholipids	
Session 10 (Week 10)	Oxidation of fatty acids, $\alpha$ -, $\beta$ - oxidation,	
Session 11 (Week 11)		
Session 12 (Week 12)	Midterm Exam	
Session 13 (Week 13)		
Session 14 (Week 14)		
Session 15 (Week 15)	Protein chemistry:	
	- Protein digestion	
	- Absorption of amino acids	
	Amino Acids definition	
	- Structure of proteins	
	- Globular Proteins	
	Fibrous Proteins	
Session 16 (Week 16)	- Urea Cycle	
	- Reactions of the urea cycle	
	- Origin of ornithine	
	Regulation of the urea cycle	
	<ul> <li>Function of the urea cycle during fasting</li> </ul>	
Session 17 (Week 17)	Protein metabolism:	
	- Nitrogen Metabolism	
	- Catabolism of Amino Group Nitrogen	
Session 18 (Week 18)	- Catabolism of Carbon Skeleton of Amino Acids	
	- Biosynthesis of Amino Acids	
Session 19 (Week 19)	-Amino Acids: Conversion to Specialized Products	
	- Disorders of Amino Acid Metabolism	
Session 20 (Week 20)	Enzymes:	
	-Properties of enzymes	
	- Mechanism of Enzyme Catalysis	
	-Michaelis–Menten kinetic theory of enzyme action	
Session 21 (Week 21)	- Regulation of enzyme activity	
	Isoenzymes	
	Enzymes in Clinical Diagnosis	
Session 22 (Week 22)	Haem metabolism:	
	- General Characteristics of Porphyrins	
	- Biological Significance of Porphyrins	
	- Synthesis of Haem	
Session 23 (Week 23)	-Disorders of Porphyrin Metabolism: Porphyria	
	- Haem Breakdown	
	- Jaundice	

Session 24 (Week 24)	Vitamins:	
	- Classification and nomenclature	
	- Sources, daily requirements	
	- Deficiency of Vitamins	
	- Water-soluble vitamins	
	- Fat-soluble vitamins	
Session 25 (Week 25)	Xenobiotic:	
	- Definition of xenobiotic	
	- Metabolism of xenobiotic	
Session 26 (Week 26)	Nucleotides and nucleic acids:	
	Nucleotides: Chemistry and Biological Significance	
	-Purine Metabolism	
	-Pyrimidine Metabolism	
Session 27 (Week 27)	-Structure of DNA	
	-Physical Properties of DNA	
	-DNA as Genetic Material	
	-Replication, translation, transcription, and repair	
Session 28 (Week 28)	-RNA Structure, Synthesis, and Processing	
	-General features of RNA	
	-Types of RNAs	
	Final theoretical Exam.	
Practical work	Practical Part:	
(one/week)	<b>1.</b> Preparation of Buffers, different types of buffer in the human body.	
	2. Qualitative test of carbohydrates.	
	3. Quantitative test of carbohydrates.	
	4. Clinical cases.	
	5. Review and practical reports	
	6. Qualitative tests of Lipids	
	7. Quantitative tests of Lipids	
	8. Clinical cases	
	9 Qualitative tests of proteins	
	10 Quantitative tests of proteins	
	Final practical exam	
Attendance	Students are expected to attend every session of class arriving on time	
Expectations	returning from breaks promptly and remaining until class is dismissed.	
F	Absences are permitted only for medical reasons and must be supported with	
	a doctor's note.	
Generic Skills	The faculty is committed to ensuring that students have the full range of	
	knowledge and skills required for full participation in all aspects of their	
	lives, including skills enabling them to be life-long learners. To ensure	
	graduates have this preparation, such generic skills as literacy and numeric,	
	computer, interpersonal communications, and critical thinking skills will be	
	embedded in all courses.	

# Pharmaceutical Microbiology I

1	Course name	Pharmaceutical Microbiology I
2	Course Code	PH207
3	Course type: /general/specialty/optional	Specialty
4	Accredited units	3 units
5	Educational hours	(2 theory + 2 Practical)
6	Pre-requisite requirements	Biology
7	Program offered the course	Department of Biomedical Science
8	Instruction Language	English language
9	Date of course approval	12/2021

Brief Description: Textbooks required for this Course:	The course will provide detailed insights into basic concepts and the differences between different classifications of microbiology. structure and function of microbes (cellular structures, metabolism, microbial growth, and its requirements and how to control it), microbial genetics, prokaryotes versus eukaryotes. This course also provides a comprehensive explanation of the microbial diversity (bacteria, viruses, fungi, and parasites) that are medically important and shows how they are defined, named, classified and reproduced, as well as of the different types of virulence factors used by pathogens to cause the diseases Microbiology an Introduction,9 <sup>th</sup> edition; (2004).Tortora GJ, Funke BR & Case CL. (Pearson International edition) Medical Microbiology, 23 <sup>rd</sup> edition; (2004). Jawetz E., Melnick JL &Adelberg's
	E. Lange Middle East edition
	Additional Resources: Lectures Notes
Course Duration	28 weeks
Delivery	<ol> <li>Lectures (Tools: board, data show).</li> <li>Practical classes (Lab experiments+ computerized experiments simulation)</li> <li>Assignments, reports and power point presentation thesis.</li> <li>Construction of illustrated posters.</li> </ol>
Course Objectives:	<ol> <li>Introduction to the nature of general microbiology and be familiar with the different branches and classification of microbiology.</li> <li>Focus on the major differences between Prokaryotes and Eukaryotes</li> <li>Know the differential identification pigments of microorganisms, understand the mechanism of action of those pigments, and identify bacteria through biochemical reactions. Plus, how to name and classify microorganisms</li> <li>Understanding the structure and morphology of bacteria and their reproduction types, growth requirements and the growth curve and metabolism</li> <li>Understanding the principles of microbial genetics and genetic study techniques</li> <li>Be familiar with the medically important Gram-negative and Grampositive pathogenic bacteria and related disease</li> <li>Focus on the medically important human pathogenic viruses and related diseases and understand their types, replication, and different classification</li> <li>Focus on the medically important human pathogenic fungi and parasites and related diseases</li> <li>In general: At the end of course the student will be able to</li> <li>Illustrate the concept of Microbiology, different microorganisms and their relation with the human diseases</li> <li>Identify the isolated microorganisms and give the medical information about.</li> <li>The student will have acquired knowledge and critical understanding of basic facts, principles and theories related to bacterial genetics.</li> </ol>

	<ul> <li>4- The student will be able to analyze unexpected results while dealing with microbes in terms of changing their characteristics and explain the abnormality at the level of microbial genetics.</li> <li>5- The student will have acquired the skill of using the microscope.</li> <li>6- The student will be able to interpret the results and find and assess the information of different microbial diseases quickly and reliably</li> <li>7- The student will be able to use the internet to search and prepare reports and do a presentation</li> </ul>	
Course Assessments	Midyear exam 20%	
	Quizzes, reports, presentation	10%
	Practical continuous assessment, exam	10%
	Final Practical exam	20%
	Final theoretical exam	40%
	Total	100%
Content Breakdown Topical Coverage	Content Breakdown Topical Coverage	
Session 1 (Week 1)	<ul> <li>Introduction to General Microbiology</li> <li>Different branches of Microbiology</li> <li>History of Microbiology</li> <li>Classification and nomenclatu</li> </ul>	blogy re of microorganisms
Session 2 (Week 2)	Study of Prokaryotes;         Classification(taxonomy) of the Prokaryotes         -       Morphology, arrangement and structure of the bacterial cell         -       The bacterial spore, The process of spore formation, Spore germination and outgrowth         Classification of bacteria, bacterial morphology and structure of bacterial cells.	
	Prokaryotic cells versus Eukaryotic Cells (structure & organelles functions)	
Session 3 (Week 3)	Bacterial nutrition and metabolism and growth requirementsPhysical Requirements • Chemical Requirements, Growth factors,Nutritional classification of microorganisms	
Session 4 (Week 4)	<b>Bacterial growth:</b> Bacterial growth curve, generation time, measurement of bacterial growth.	
Session 5 (Week 5)	<ul> <li>Bacterial growth control:</li> <li>Antiseptic versus disinfectants (types &amp; mechanisms)</li> <li>Sterilization methods: <ul> <li>Heat sterilization (dry, moist)</li> <li>Cold sterilization (radiation, gas, filtration).</li> </ul> </li> </ul>	
Session 6 (Week 6)	Different bacterial stains:	
	<ul><li> (Gram stain, Acid-fast satin</li><li> Principles and reactions</li></ul>	etc)

	Microbial biochemical reactions	
Session 7 (Week 7)	Different types of culture media (constituents and characteristics)	
	(Enrichment growth media, differential media and selective media)	
	- Bacterial biochemical reaction	
Session 8 (Week 8)	All medical important human pathogenic Gram-positive Staphylococci	
	bacteria and related disease with virulence factor	
Session 9 (Week 9)	All medical important human pathogenic Gram-positive Streptococci	
Eastion 10 (Weak 10)	Discretia and related disease with virulence factor	
Session 10 (Week 10)	Clostridium, Corynebacterium, Bacilius, Enterococcus, and Pheumococcus	
Session 12 (Week 11)		
Session 12 (Week 12)	Midvear exam	
Session 13 (Week 13)		
Session 14 (Week 14)		
Session 15 (Week 15)	Medical Important numan pathogenic Gram-negative	
	Litteropacteriaceaebacteria and related disease with withence factor	
Session 16 (Week 16)	Continue to Medical important human pathogenic Gram-negative bacteria	
. ,	and related disease with virulence factor	
	(Pseudomonas, Vibrio, Campylobacter and Helicobacter, Brucella, Yersinia,	
	Acinetobacterbaumannii, etc)	
Session 17 (Week 17)	Acid fast bacteria: Mycobacteria species with related diseases and virulence	
	factor	
Session 18 (Week 18)	Unusual bacteria and related diseases with virulence factor	
	<ul> <li>-Spirochetes (Treponema, Borrelia.)</li> </ul>	
	-Moraxella	
	<ul> <li>-Mycoplasma, Ricketteslae.</li> <li>-Chlamydia</li> </ul>	
Session 10 (Week 10)	• -Childhydda Werkshen (Assignment, discussion and presentation)	
Session 20 (Week 19)	Microbial Constice:	
JESSION 20 (WEEK 20)	a. Nucleic acid types. Structure and Function (DNA & RNA)	
	b. Differences between DNA & RNA	
	c. Protein synthesis (Replication, transcription & translation)	
Session 21 (Week 21)	Continue to Microbial Genetics.	
	a. Changes in Genetic Material (Mutation • Types of	
	Mutations • Mutagens)	
Session 22 (Mark 22)	Continue to Microbial Genetics	
JESSION 22 (WEEK 22)	a. Genetic Transfer and Recombination (Plasmids and Transposons	
	Transformation in Bacteria      Conjugation in Bacteria	
	Transduction in Bacteria, Genetic Mechanisms of Drug	
	Resistance in Bacteria)	
	<ul> <li>Blotting Techniques • Polymerase Chain Reaction (PCR) • Gene Thoracut</li> </ul>	
Section 22 (Mark 22)	Study of Virology	
Session 23 (Week 23)	Study of Virology:	
	Types, classification, structures characteristics & replication of viruses	
Session 24 (Week 24)	Continue to virology:	

	Medically important human pathogenic viruses and related diseases	
Section 2E (Meak 2E)	Study of Mycology:	
Session 25 (Week 25)	Suuy of Mycology:	
	Types, classification, structures characteristics & replication of fungi	
Session 26 (Week 26)	Continue to Mycology:	
	Medically important human pathogenic fungi and related diseases	
Session 27 (Week 27)	Study of parasites:	
	common classification, and characteristics of parasites	
Session 28 (Week 28)	Continue to parasites:	
	Medically important human pathogenic fungi and related diseases .	
	Final theoretical Exam	
Practical work	1-Laboratory regulations: -Equipment and aids in microbiology labse.g.	
(one/week)	incubator, laminar flow, aseptic hood, autoclave, hot air sterilizer, deep	
	freezer, refrigerator,	
	2-Use of microscope: -Examination of stained smears. Examination of wet	
	preparations.	
	3. Sterilization of glassware, preparation and sterilization of media.	
	4-Culture media: -Basic, enriched, selective and differential media. Agars	
	and broth. Aerobic and anderobic cultures	
	6-Microbiological status I: Preparation of smears, Simple status Negative	
	stain	
	7-Microbiological stains II: -Gram stain, Acid fast, capsule and spore stain.	
	8-Systematic bacteriology: -Morphology and staining of microorganisms of	
	medical importanceCulture characteristicsBiochemical tests.	
	9. Microbiological assay of antibiotics by cup plate method and other	
	methods	
	10. Motility determination by Hanging drop method.	
	11. Sterility testing of pharmaceuticals.	
	Final Practical Exam	
Attendance	Students are expected to attend every session of class, arriving on time,	
Expectations	returning from breaks promptly and remaining until class is dismissed.	
	Absences are permitted only for medical reasons and must be supported	
	with a doctor's note.	
Generic Skills	By the end of the course the students will have the full range of knowledge	
	and skills required for full participation in all aspects of their lives, including	
	skills enabling them to be life-long learners. To ensure graduates have this	
	preparation, such generic skills as literacy and numeric, computer,	
	interpersonal communications, and critical thinking skills will be embedded	
	in this course	
Course Change	Content of the courses is revised on an ongoing basis to ensure relevance to	
course enunge	changing educational employment and marketing needs. The instructor will	
	endeavor to provide notice of changes to students if hannen. Timetables	
	also will be revised continuously	

#### **Clinical biochemistry**

1	Course name	Clinical Biochemistry
2	Course Code	РН 304
3	Course type: /general/specialty/optional	General
4	Accredited units	3 units (2 hrs./week theoretical
		2 hrs./week practical)
5	Educational hours	4 hours / week
6	Pre-requisite requirements	Biology, pathology and organic chemistry
7	Program offered the course	Department of Pharmaceutical chemistry
8	Instruction Language	English
9	Date of course approval	12/2021

Brief Description:	Clinical biochemistry is the division c	of laboratory medicine that deals with
Ditter 2 courter to the	the measurement of chemicals (both	natural and unnatural) in blood, urine
	and other body fluids. The subject de	als with study of acid-base balance of
	the body, normal and abnormal serum levels of electrolytes, enzyme,	
	hormones and CBC levels.	
Textbooks required	1- Clinical biochemistry. By GawMurpl	ny cowanetal. 4 <sup>th</sup> edition. Elsiver
for this Csourse:	2. Clinical Biochemistry: An illustrate	ed color text. By Allan Gaw, Michael J
	Murphy, Rajeev Srivastava, Robert A Cowan, and Denis St J O'Reilly. Elsevier	
	Ltd	
	3. Basic Concepts in Clinical Biochemistry: A Practical Guide. By Vijay	
	Kumar, Kiran Dip Gill. Springer Nature.	
	4. Clinical Biochemistry: Lecture Note	S. By -Peter Rae, Mikeurane, Repecca
Course Duration	Pattenden. John Wiley & Sons Ltu	
Course Duration	28 Weeks	
Delivery	• Lecture-based, Group interaction and discussion, Use of video technique,	
Course Objectives	practical classes.	
Course Objectives:	By the end of the biochemical compositi	Juid be able to:
	2 Summarizing changes in biochemica	al indicators of diseases
	2. Summarizing changes in diochemical indicators of diseases	
	with different disease states.	
	4. Explanation of ways to ensure	the quality of medical laboratories.
	performance and the interpretation o	f the various results.
	5. Determining the accurate descript	ion of changes in blood chemisty and
	describing the disease and its origin.	
Course Assessments		200/
Course Assessments		20%
	Quizzes, reports, presentation	10%
	Practical continuous assessment,	10%
	Final Practical evam	20%
	Final theoretical exam	10%
	Total	100%
Content Breakdown	Content Breakdown Tonical Covers	100%
Topical Coverage	Content Dicakuown Topicar Covera	.ge
Session 1(Week 1)	Unit I: Introduction: (3 hr.)	
	a. Definition of clinical biochemistry	laboratory
	b. The use of the laboratory	-
	c. The interpretation of results	
	d. Reference intervals	
	e. Specimen collection	
Session 2(Week 2)	Unit 2-Fluid, electrolyte balance adre	nal functions (9hrs.)
	a. Concept and vocabulary	
	b. types of buttering system.	
Genter 2(West 2)		
Session 5(week 5)	u. пуреглаtremia and hyponatremia	
Contar (Weak 1)	e .Hyperkalerina and hypokalerina	- 10 km
Session 4(week 4)	Unit 3. Investigation of renal function	1: (6 nr)
	a. Tunctions of the kidney	

	b. Glomerular function	
	c. Principal of GFR	
	d. Renal tubular function	
Session 5(Week 5)	e. Acute renal failure	
	t. Chronic renal failure	
	t. Iubular dysfunction	
	g. Specific proteinuria	
Section (Wealt ()	II. Glycosulla	
Session o(week o)	Acid hase metabolic disorders	
	b. Blood gases and respiratory disorders	
Session 7(Week 7)	c. plasma protoin and enzymos of clinical significance	
Session / (Week /)	d. Immunoglobulin and acute phase proteins	
Session 8(Week 8)	Unit 5 Endocrinology and diabetes (15 hrs.)	
Session o(Week o)	a Biochemical regulators	
	b. Hormone structure	
	c. Assessment of endocrine Control	
	d. Types of endocrine control	
Session 9(Week 9)	a. Pituitary function (Anterior & Posterior pituitary hormones	
	b. Pituitary tumors	
	c. Hypopituitarism	
	d. Growth disorders and acromegaly	
	e. Growth hormone insufficiency	
Session 10(Week 10)	a. Thyroid biochemistry and thyroid disorders (- Goiter, Grave's disease,	
	hypothyroidism, hyperthyroidism).	
	b.Adrenal biochemistry and adrenal disorders (Hypofunctionand	
	Hyperfunction of the adrenal cortex)	
Session 11(Week 11)		
Session 12(Week 12)	Midvear Fxam	
Session 13(Week 13)		
Session 14(Week 14)		
Session 16(Week 16)	a. Gonadal functions and dynamic functions tests	
	b. g. Hypothalamic–pituitary–gonadal axis	
	c. h. Disorders of male sex hormones	
	d. Disorders of female sex hormones	
Session 17(Week 17)	Glucose metabolism and diabetes mellitus:	
	a. Insulin b. Diabatas mallitus	
	D. Diabeles memilius	
	d Diagnosis and monitoring of diabetes mellitus	
Session 18(Week 18)	Linit 6 GIT biochemistry (3 br)	
Session 10(Week 10)	a Enzyme analysis	
	b. Fecal analysis	
Session 19(Week 19)	Unit 7. Plasma enzyme in diagnosis: (3 hr)	
	Introduction, assays of enzymes, and enzymes of diagnostic value	
Session 20(Week 20)	Unit 8. Clinical biochemistry of the Liver ( 6 hrs. )	
	a. Liver function tests and liver enzyme	
	b. bilirubin metabolism	
	c.Jaundice and liver diseases: acute and chronic liver diseases.	

Session 21(week 21)	c.Viral Hepatitis: investigation and interpretation of lab findings	
Session 22(Week 22)	Unit 9. Plasma lipid & Lipoprotein: (3 hr)	
	a. Chemical classification of plasma lipid & atherogenic profiles.	
	b. Classification of plasma lipoproteins, metabolism, & their function	
Session 23(Week 23)	Unit 10-Clinical biochemistry of CVS( 3 hrs. )	
	a. Myocardial infarction.	
	b. Cardiac markers	
	c. Hypertension	
Session 24(Week 24)	Unit 11. Clinical biochemistry of skeleton( 3 hrs. )	
	a.Calcium regulation, calcium status and bone metabolism	
	b. Bone diseases, osteoporosis and rickets	
	c. Hyperuricemia and gout	
	d. Risk factors.	
Session 25(Week 25)	Unit 12. Hematology: (3 hr)	
	a. Red & white blood cells)	
	<ul> <li>Blood cells production( site of production, hematopoiesis)</li> </ul>	
	c. Anemias	
	d. leukemias	
Session 26(Week 26)	Unit 13. Special topics ( 9 hrs. )	
	a.Paediatric biochemistry	
	biochemistry of Pregnancy and Clinical Gynecology	
	c.Point of care testing ( POCT)	
Session 27(Week 27)	d.DNA diagnosis	
	e.Tumor markers	
Session 28(Week 28)	f.Therapeutic drug monitoring	
	g.Fetal monitoring, prenatal diagnosis and Newborn screenin	
	Final theoretical exam.	
Practical work	Practical Part:	
(one/week)	1. Common Clinical Laboratory Hazards and Waste Disposal	
	2 Blood Collection	
	3-Quality Control in Laboratory	
	<ul><li>3-Quality Control in Laboratory</li><li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li></ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Discharging tests in lines function. Case history</li> </ul>	
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	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types, Laboratory investigation of iron disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders or bong disease</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types, Laboratory investigation firon disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders orbone disease</li> <li>15. Dynamic function tests (Insulin stress test, GnRH test, OGTT with GH</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types, Laboratory investigation firon disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders orbone disease</li> <li>15. Dynamic function tests (Insulin stress test, GnRH test, OGTT with GH, Measurement TRH test )</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types,Laboratory investigationof iron disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders orbone disease</li> <li>15. Dynamic function tests (Insulin stress test, GnRH test, OGTT with GH, Measurement, TRH test,)</li> <li>16. Pituitary function. Case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types,Laboratory investigationof iron disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders orbone disease</li> <li>15. Dynamic function tests (Insulin stress test, GnRH test, OGTT with GH, Measurement, TRH test,)</li> <li>16. Pituitary function, Case history</li> <li>17. Tests of growth hormone Insufficiency -Case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types, Laboratory investigation firon disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders orbone disease</li> <li>15. Dynamic function tests (Insulin stress test, GnRH test, OGTT with GH, Measurement, TRH test,)</li> <li>16. Pituitary function, Case history</li> <li>17. Tests of growth hormone Insufficiency, -Case history</li> <li>18. Thyroid function tests, -Case history</li> </ul>	
	<ul> <li>3-Quality Control in Laboratory</li> <li>4. Determination of blood acidity and alkalinity (blood gases). Case history</li> <li>5. Investigation&amp; case history</li> <li>6. Estimation of Urea in Serum and Urine</li> <li>7. Determine Urea Clearance</li> <li>8Estimated GFR. Case history</li> <li>9. Review</li> <li>10. Biochemical tests in liver function. Case history</li> <li>11.RBS, FBS, OGTT. Case history</li> <li>12. Lipid Profile in Serum Sample. Case history</li> <li>13. CBC, blood types,Laboratory investigationof iron disorders, Case history</li> <li>14. Biochemistry testing in calcium disorders orbone disease</li> <li>15. Dynamic function tests (Insulin stress test, GnRH test, OGTT with GH, Measurement, TRH test,)</li> <li>16. Pituitary function, Case history</li> <li>17. Tests of growth hormone Insufficiency, -Case history</li> <li>18. Thyroid function testsCase history</li> <li>19. Assessing the Hypothalamic-Pituitary-adrenocortical. Axis function</li> </ul>	

	<ul><li>20Synacthen tests – Dexamethasone, - suppression tests, - Case history.</li><li>21. Urine analysis and fecal analysis</li></ul>	
	Final practical Exam	
Attendance Expectations	Students are expected to attend every session of class, arriving on time, returning from breaks promptly and remaining until class is dismissed. Absences are permitted only for medical reasons and must be supported with a doctor's note.	
Generic Skills	The faculty is committed to ensuring that students have the full range of knowledge and skills required for full participation in all aspects of their lives, including skills enabling them to be life-long learners. To ensure graduates have this preparation, such generic skills as literacy and numeric, computer, interpersonal communications, and critical thinking skills will be embedded in all courses.	

### Pharmaceutical Microbiology II

1	Course name	Pharmaceutical Microbiology II
2	Course Code	РН307
3	Course type: /general/specialty/optional	Specialty
4	Accredited units	3 units (3 theory + 2 Practical)
5	Educational hours	5 hours/week
6	Pre-requisite requirements	Microbiology II
7	Program offered the course	Bachelor Degree in Pharmaceutical Sciences
8	Instruction Language	English language
9	Date of course approval	12/2021

Brief Description:	This course provides learning opportunities for understanding the principles of immunology and being familiar with the role of the immune system in health well-being. In addition, this course is taught pathogenicity (the mechanisms by which the microbes cause the diseases). Also, this course explains the phenomena of biofilm and the mechanisms by which the pathogens resist antimicrobial agents, and how the antimicrobial agents fight the pathogens. Furthermore, this course is concerned with providing knowledge about how to explore new antimicrobial agents from microbes to kill other microbes by use of fermentation and recombinant DNA techniques	
Textbooks required for this Course:	<ol> <li>Pharmaceutical biotechnology Fundamental and application, 5<sup>th</sup> Edition. Daan J.A. Crommelin, Robert D. Sindelar, Bernd Meibohm Editors. Springer</li> <li>Pharmaceutical biotechnology Concept and application. Gary Walsh. John Welly&amp;Sons.Ltd</li> <li>Microbiology an Introduction,9<sup>th</sup> edition; (2004).Tortora GJ, Funke BR &amp; Case CL. (Pearson International edition)</li> <li>Additional Resources: Lectures Notes</li> </ol>	
Course Duration	28 weeks	
Delivery	<ol> <li>Lectures (Tools: board, data show).</li> <li>Practical classes (Lab experiments+ computerized experiments simulation)</li> <li>Assignments, reports and power point presentation thesis.</li> <li>Construction of illustrated posters.</li> </ol>	

Course Objectives:	At the end of course the student will be able to		
	1. explain the nature and definitions of immunology, illustrate the concept of		
	immunology and its function and de	scribe the differences between the	
	innate and the acquired immune system.		
	2. The student will be able to explain the	ne microbial mechanism by which	
	microbes cause disease and to expla	in the properties of biofilms and their	
	The student will be able to explain the	a concent and importance of industrial	
	pharmaceutical microbiology.		
	4. The student will be able to explain the basics of fermentation techniques		
	and their importance in exploring new pharmaceutical substances of		
	interest.		
	5. The student will be able to demonstrate the technique of cloning and gene		
	expression and its importance in the exploration of new pharmaceutical		
	materials of interest.		
	<ol> <li>The student will he able to interpret</li> </ol>	the results and find and assess the	
	information of different microbial di	seases quickly and reliably.	
	8. The student will be able to use the ir	iternet to search and prepare reports	
	and do a presentation.		
Course Assessments	Midyear exam	20%	
	Quizzes, reports, presentation	10%	
	Practical continuous assessment,	10%	
	exam		
	Final Practical exam	20%	
	Final theoretical exam	40%	
	Total	100%	
Content Breakdown	Content Breakdown Topical Coverage		
Topical Coverage			
Session 1 (Week 1)	General introduction to immunology		
Session 2 (Week 2)	Innate immune system (cells, functions and mechanisms)		
Session 3 (Week 3)	Acquired immune system(cells, function	s and mechanisms)	
Session 4 (Week 4)	Autoimmune diseases and Hypersensitivity		
Session 5 (Week 5)	Workshop session / scientific activity/ presentation		
Session 6 (Week 6)	Introduction in microbial Pathogenicity &	& predispose factors of infection with	
	True and Opportunistic pathogens		
Session 7 (Week 7)	Microbial Portal of entry and portal of experience for the	kit and specificity of portals to specific	
	Major steps of nathogenicity & minimum	n infectious dose (MID) & Strategies of	
	microbe to evade the host defense		
Session 8 (Week 8)	Classification of antimicrobial agents according to the microbial biology		
	Mode of action of antibiotics against pat	hogenic bacteria, bacteriostatic and	
	bactericide concept and		
Session 9 (Week 9)	Assessment of minimum inhibitory conce	entration (MIC) and minimum	
	bactericidal concentration (MBC)		
Session 10 (Week 10)	Workshop session / scientific activity/ presentation		
Session 11 (Week 11)			

Session 12 (Week 12)	
Session 13 (Week 13)	Midyear exam
Session 14 (Week 14)	
Session 15 (Week 15)	Strategies and predispose factors of bacterial resistance. (Miss use & over use of antibiotics, capsules, cell wall components, antigenic
Cossien 1C (Mask 1C)	Variation)
Session 16 (week 16)	Quorum Sensing and dispersal
Session 17 (Week 17)	Antibiofilm agents (natural and synthetics)
Session 18 (Week 18)	Biofilm risk on medicine and industry and types of anti-biofilm agents (natural and chemical)
Session 19 (Week 19)	Rate of Microbial Death and Conditions influencing the Effectiveness of Antimicrobial Agent Activity.
Session 20 (Week 20)	Workshop (Assignment, discussion and presentation)
Session 21 (Week 21)	History, medical importance and general introduction to pharmaceutical biotechnology (Fermentation and type of fermentation & Recombinant DNA) Fermentation: (fermenter, Media, Batch/continuous/feed batch fermentation) and differences between fermenter and Bioreactor
Session 22 (Week 22)	Fermentation Production ofvinegar (slow process, quick process, and natural fermentation and uses of vinegar), and alcohol
Session 23 (Week 23)	Fermentation Production of vitamins and amino acids
Session 24 (Week 24)	Fermentation Production of organic acid; Lactic acid
Session 25 (Week 25)	Fermentation Production of organic acid; Acetic acid
Session 26 (Week 26)	Fermentation production of antibiotics, steroid s
Session 27 (Week 27)	Production of vaccines from microbes
Session 28 (Week 28)	Gen cloning (recombinant DNA)
	Final theoretical exam
Practical work (one/week)	<ul> <li>1-Laboratory regulations: -Equipment and aids in microbiology labs.</li> <li>2-Determination of minimum inhibitory concentration(MIC)</li> <li>3- Determination of minimum bactericidal concentration (MBC)</li> <li>4-Assessment of bacterial sensitivity to antibiotics.</li> <li>5- Assessment of bacterial resistance to antibiotics.</li> <li>6- Practicing the fermentation technique</li> </ul>
	Final practical exam
Attendance Expectations	Students are expected to attend every session of class, arriving on time, returning from breaks promptly and remaining until class is dismissed. Absences are permitted only for medical reasons and must be supported with a doctor's note.
Generic Skills	By the end of the course the students will have the full range of knowledge and skills required for full participation in all aspects of their lives, including skills enabling them to be life-long learners. To ensure graduates have this preparation, such generic skills as literacy and numeric, computer, interpersonal communications, and critical thinking skills will be embedded in this course
Course Change	Content of the courses is revised on an ongoing basis to ensure relevance to

changing educational employment and marketing needs. The instructor will
endeavor to provide notice of changes to students if happen. Timetables also
will be revised continuously.

# Pharmaceutical Biotechnology

1	Course name	Pharmaceutical biotechnology
2	Course Code	PH 401
3	Course type: /general/specialty/optional	2 hours theory
4	Accredited units	2 units
5	Educational hours	2 hours/week
6	Pre-requisite requirements	Biology
7	Program offered the course	Department of biomedical science
8	Instruction Language	English
9	Date of course approval	12/2021

Brief Description:	Biotechnology has a long promise	to revolutionize the biological
	sciences and technology.	
	<ul> <li>Scientific application of biotechno</li> </ul>	logy in the field of genetic
	engineering, medicine and fermer	itation technologymakes the subject
	Interesting.	
	<ul> <li>Biotechnology is leading to new bio provention and cure of diseases.</li> </ul>	ological revolutions in diagnosis,
	drugs	
	<ul> <li>Biotechnology has already product</li> </ul>	ed transgenic crops and animals and
	the future promises lot more.	
	It is basically a research-based subject.	
Textbooks required	1. B.R. Glick and J.J. Pasternak: Molecular Biotechnology: Principles and	
for this Course:	Applications of RecombinantDNA: ASM Press Washington D.C.	
	2. RA Goldshy et. al., : Kuby Immunolo	gy.
	3. J.W. Goding: Monoclonal Antibodie	S.
	4. J.M. Walker and E.B. Gingold: Molec	cular Biology and Biotechnology by
	5. Zaborsky: Immobilized Enzymes. CR	C Press, Degraland, Ohio
	6. S.B. Primrose: Molecular Biotechno	ogy (Second Edition) Blackwell
	Scientific Publication.	
	7. Stanbury F., P., Whitakar A., and Ha	ll J., S., Principles of fermentation
	technology, 2nd edition, Aditya books	Ltd., New Delhi
Course Duration	28 weeks	
Deliverv	<ul> <li>Lecture-based, Group interaction ar</li> </ul>	nd discussion, Use of video technique,
,	practical classes.	
Course Objectives	Upon completion of the subject student shall be able to;	
course objectives:		
Course Objectives.	1. Understanding the importance of In	nmobilized enzymes in
Course Objectives.	1. Understanding the importance of In Pharmaceutical	nmobilized enzymes in
Course Objectives.	<ol> <li>Understanding the importance of In Pharmaceutical Industries</li> <li>Genetic engineering applications in</li> </ol>	nmobilized enzymes in relation to production of
Course Objectives.	<ol> <li>Understanding the importance of In Pharmaceutical Industries</li> <li>Genetic engineering applications in pharmaceuticals</li> </ol>	nmobilized enzymes in relation to production of
Course Objectives.	<ol> <li>Understanding the importance of In Pharmaceutical Industries</li> <li>Genetic engineering applications in pharmaceuticals</li> <li>Importance of Monoclonal antibodi</li> </ol>	nmobilized enzymes in relation to production of es in Industries
Course Objectives.	<ol> <li>Understanding the importance of In Pharmaceutical Industries</li> <li>Genetic engineering applications in pharmaceuticals</li> <li>Importance of Monoclonal antibodi</li> <li>Appreciate the use of microorganism</li> </ol>	nmobilized enzymes in relation to production of es in Industries ms in fermentation technology
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Course Assessments	<ol> <li>Understanding the importance of In Pharmaceutical Industries</li> <li>Genetic engineering applications in pharmaceuticals</li> <li>Importance of Monoclonal antibodi</li> <li>Appreciate the use of microorganism</li> <li>Midyear exam</li> </ol>	relation to production of es in Industries ms in fermentation technology
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Course Assessments	<ol> <li>Understanding the importance of In Pharmaceutical Industries</li> <li>Genetic engineering applications in pharmaceuticals</li> <li>Importance of Monoclonal antibodi</li> <li>Appreciate the use of microorganism</li> <li>Midyear exam</li> <li>Quizzes, reports, presentation</li> <li>Practical continuous assessment, exam</li> </ol>	relation to production of es in Industries ms in fermentation technology 20% 10%
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Course Assessments Course Assessments Content Breakdown Topical Coverage Session 1 (Week 1)	1. Understanding the importance of In         Pharmaceutical         Industries         2. Genetic engineering applications in         pharmaceuticals         3. Importance of Monoclonal antibodi         4. Appreciate the use of microorganism         Midyear exam         Quizzes, reports, presentation         Practical continuous assessment,         exam         Final Practical exam         Final theoretical exam         Total         Content Breakdown Topical Coverage         Unit I       (10 Hours)	Introduction of         relation to production of         es in Industries         ms in fermentation technology         20%         10%         20%         10%         10%         20%         40%         100%
Course Assessments Content Breakdown Topical Coverage Session 1 (Week 1)	1. Understanding the importance of In         Pharmaceutical         Industries         2. Genetic engineering applications in         pharmaceuticals         3. Importance of Monoclonal antibodi         4. Appreciate the use of microorganism         Midyear exam         Quizzes, reports, presentation         Practical continuous assessment,         exam         Final Practical exam         Final theoretical exam         Total         Content Breakdown Topical Coverage         Unit I       (10 Hours)         a) Brief introduction to Biotechnology	<pre>int shall be use to; nmobilized enzymes in relation to production of es in Industries ms in fermentation technology 20% 20% 20% 40% 10% 20% 40% 100% </pre>
Course Assessments Content Breakdown Topical Coverage Session 1 (Week 1)	1. Understanding the importance of In         Pharmaceutical         Industries         2. Genetic engineering applications in         pharmaceuticals         3. Importance of Monoclonal antibodi         4. Appreciate the use of microorganism         Midyear exam         Quizzes, reports, presentation         Practical continuous assessment,         exam         Final Practical exam         Final theoretical exam         Total         Content Breakdown Topical Coverage         Unit I       (10 Hours)         a) Brief introduction to Biotechnology         Sciences.	Introduction of         relation to production of         es in Industries         ms in fermentation technology         20%         10%         20%         40%         100%         with reference to Pharmaceutical
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Course Assessments Content Breakdown Topical Coverage Session 1 (Week 1)	1. Understanding the importance of In         Pharmaceutical         Industries         2. Genetic engineering applications in         pharmaceuticals         3. Importance of Monoclonal antibodi         4. Appreciate the use of microorganism         Midyear exam         Quizzes, reports, presentation         Practical continuous assessment,         exam         Final Practical exam         Final theoretical exam         Total         Content Breakdown Topical Coverage         Unit I       (10 Hours)         a) Brief introduction to Biotechnology         Sciences.         b) Enzyme Biotechnology- Methods of applications.	Introduction of         relation to production of         es in Industries         ms in fermentation technology         20%         10%         20%         10%         20%         40%         100%         set of Pharmaceutical         enzyme immobilization and

	Industries.	
	d) Brief introduction to Protein Engineering.	
	e) Use of microbes in industry. Production of Enzymes- General	
	consideration -Amylase, Catalase, Peroxidase, Lipase, Protease,	
	Penicillinase.	
Session 3 (Week 3)	f) Basic principles of genetic engineering.	
Session 4 (Week 4)	g) Brief introduction to PCR.	
Session 5 (Week 5)	<ul> <li>h) Study of cloning vectors, restriction endonucleases and DNA ligase.</li> <li>i) Overview protein and protein chemistry.</li> </ul>	
Session 6 (Week 6)	j) Protein extraction and protein purification (chromatography)	
Session 7 (Week 7)	k) protein assay and electrophoresis (agarose and SDS-PAGE)	
Session 8 (Week 8)	Unit II (10 hours)	
	a. Florescence protein, FRET and using fluorescence in measurement drug protein interaction.	
Session 9 (Week 9)	b) Recombinant protein	
Session 10 (Week 10)	c) Recombinant DNA technology. Application of genetic engineering in medicine.	
	c) Application of r DNA technology and genetic engineering in the	
	production of:	
	i) Interferon	
	ii) Vaccines- hepatitis- B	
Session 11 (Week 11)		
Session 12 (Week 12)	Miducar over	
Session 13 (Week 13)	Midyear exam	
Session 14 (Week 14)		
Session 15 (Week 15)	iii) Hormones-Insulin.	
Session 16 (Week 16)	d) Brief introduction to PCR	
Session 17 (Week 17)	Unit III (10 Hours)	
	Types of immunity- humoral immunity, cellular immunity	
	a) Structure of Immunoglobulins	
	b) Structure and Function of MHC	
Session 18 (Week 18)	c) Hypersensitivity reactions, Immune stimulation and Immune	
	suppressions.	
	vaccine antitoxins, serum-immune blood derivatives and other products	
	relative to immunity	
Session 19 (Week 19)	e) Storage conditions and stability of official vaccines	
Session 20 (Week 20)	f) Hybridoma technology- Production, Purification and Applications	
	g) Blood products and Plasma Substituties	
Session 21 (Week 21)	Unit IV (08Hours)	
	a) Immuno blotting techniques- ELISA, Western blotting, Southern blotting.	
	b) Genetic organization of Eukaryotes and Prokaryotes	
Session 22 (Week 22)	c) Microbial genetics including transformation, transduction, conjugation,	
	I plasmids and transposons	
Session 23 (Week 23)	d) Introduction to Microbial biotransformation and applications.	

Session 25 (Week 25)	Unit V (07 Hours)
	a) Fermentation methods and general requirements, study of media,
	equipments, sterilization methods, aeration process, stirring.
Session 26 (Week 26)	b) Large scale production fermenter design and its various controls.
	c) Study of the production of - penicillins, citric acid, Vitamin B12, Glutamic
	acid, Griseofulvin,
Session 27 (Week 27)	c) Study of the production of - penicillins, citric acid, Vitamin B12, Glutamic
	acid, Griseofulvin,
Session 28 (Week 28)	d) Blood Products: Collection, Processing and Storage of whole human
	blood, driedhuman plasma, plasma Substitutes.
	Final theoretical Exam.
Practical work	1. Competent bacteria (Top 10 and BL21).
(one/week)	2. Insert plasmid in competent bacteria.
	3. protein production and purification (affinity column).
	4. Assay protein.
	Final Practical Exam
Attendance	Students are expected to attend every session of class, arriving on time,
Expectations	returning from breaks promptly and remaining until class is dismissed.
	Absences are permitted only for medical reasons and must be supported
	with a doctor's note.
Generic Skills	The faculty is committed to ensuring that students have the full range of
	knowledge and skills required for full participation in all aspects of their
	lives, including skills enabling them to be life-long learners. To ensure
	graduates have this preparation, such generic skills as literacy and numeric,
	computer, interpersonal communications, and critical thinking skills will be
	embedded in all courses.